

Inflammatory response of peri-implant cells to piezoelectric-ceramic composite implant surfaces – *in vitro* study

Fernandes, B.¹, Tainen, L.², Gasik, M.³, Carvalho, O.², Cruz, M.¹, Mata, A.^{4,5}

¹ Grupo de Investigação em Biologia e Bioquímica Oraís (GIBBO-UICOB), Faculdade de Medicina Dentária, Universidade de Lisboa, Lisboa, Portugal;

² Center for Microelectromechanical Systems (CMEMS), Departamento de Engenharia Mecânica, Universidade do Minho, Guimarães, Portugal

³ Department of Chemical and Metallurgical Engineering, Aalto University, Espoo, Finlândia

⁴ Grupo de Investigação em Biologia e Bioquímica Oraís – LIBPhys FCT UID/FIS/04559/2013, Faculdade de Medicina Dentária, Universidade de Lisboa, Lisboa, Portugal

⁵ CEMDBE – Cochrane Portugal, Faculdade de Medicina Dentária, Universidade de Lisboa, Lisboa, Portugal

BACKGROUND AND OBJETIVE

Osseointegration is the major success factor for dental implants.¹ In order to optimize the biological response, several strategies have been investigated.² Due to its piezoelectric properties similar to bone electric potentials generated in loading function, barium titanate (BaTiO₃) piezoelectric ceramic is a potential approach for promoting osteogenic proliferation and differentiation and, consequently, accelerating osseointegration.^{3,4} In this sense, a new technique was developed to stimulate the cells in the implant bed through the use of piezoelectric ceramics as a source of electrical stimulation. However, the potential inflammatory effects in peri-implant cells and tissues are not defined.

The aim of this *in vitro* study was to evaluate the inflammatory response of gingival fibroblasts and human fetal osteoblasts in contact with barium titanate functionalized zirconia implant surfaces with piezoelectric properties.

MATERIALS AND METHODS

Composite discs with 5 wt.% BaTiO₃ in Yttria-stabilized zirconia (YSZ) were prepared through press-and-sintering technique (n=15). Contact poling was carried out in silicon oil bath under DC 2 kV/mm electric field at 130 °C for 30 min followed with field cooling.⁶ Reference samples of YSZ were processed parallel to the composites. Human gingival fibroblasts (hTERT) and fetal osteoblasts (hFOB1.19) were cultured on discs for 14 and 7 days, respectively. Cell viability was evaluated at 1, 3, 7 and 14 days using a commercial resazurin-based method. IL-1β and IL-6 were evaluated at 1 and 3 days in each fibroblast and osteoblast culture and osteopontin was measured in osteoblast cell culture at 3- and 7-days using ELISA (pg/mL). Osteoblasts alkaline phosphatase (ALP) activity was measured using an enzymatic colorimetric assay at 7 and 14 days (μmol/min/mL). All results were presented as mean ± confidence interval (CI). Group comparisons were based on one-way ANOVA repeated measures or Kruskal-Wallis and Tukey's post-hoc using appropriate statistical software (IBM® SPSS® for Mac version 27.0.1.0) and significance was set at p<0.05.

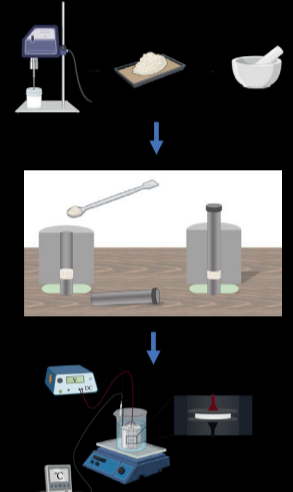


Figure 1 – Illustrative images of sample preparation. Image created with BioRender ©.

RESULTS

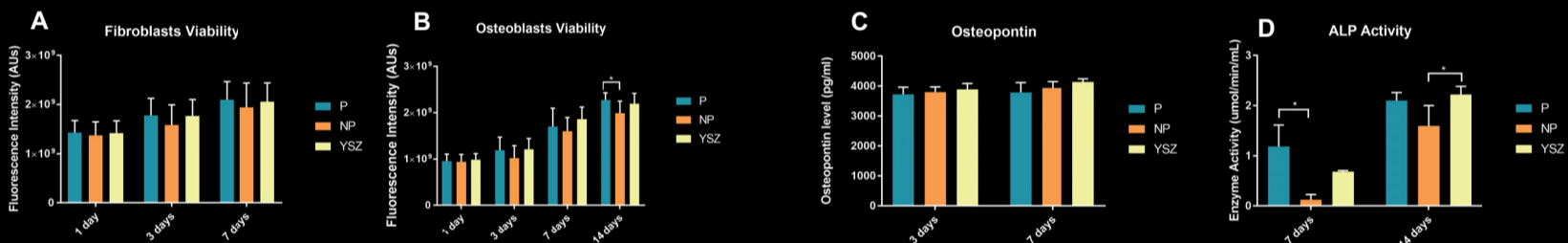


Figure 1 – Bar chart with mean and standard deviation of fibroblasts (A) and osteoblasts viability (B) using fluorescence intensity expressed in arbitrary units – AU. Statistical significance: *p<0.05.

Figure 2 – Bar charts showing osteopontin levels (C) and ALP activity (D) as mean concentration in pg/mL and enzymatic activity in μmol/min/mL, respectively. Error bars represent standard deviation. One-way repeated measures ANOVA with post-hoc Tukey test were used for comparisons between study groups. Statistical significance: *p<0.05.

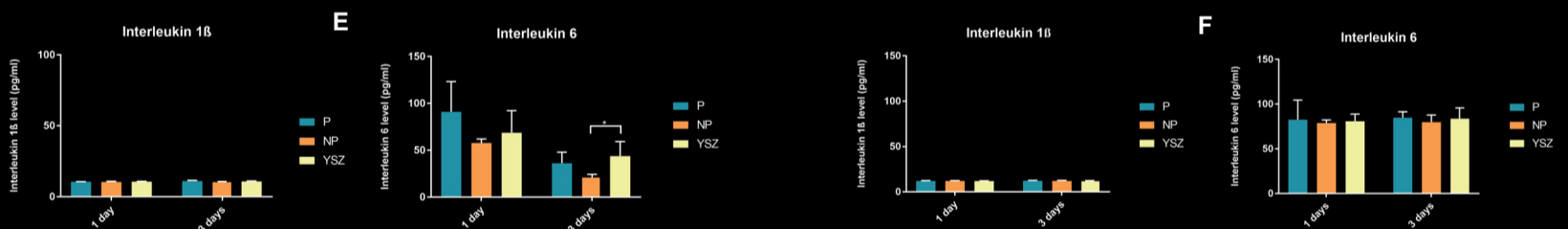


Figure 3 – Bar charts showing fibroblasts (E) and osteoblasts (F) interleukin 1β and interleukin 6 levels as mean concentration in pg/mL. Error bars represent standard deviation. One-way repeated measures ANOVA with post-hoc Tukey test were used for comparisons between study groups. Statistical significance: *p<0.05.

DISCUSSION

Poled zirconia surfaces with 5% BaTiO₃ demonstrated superior results of osteoblasts viability and initial differentiation compared to non-poled samples. These results agree with the literature.³⁻⁵ Fibroblasts behavior does not seem to be influenced by polarization. Concerning osteoblasts inflammatory markers, IL-1β secretion remain constant and an increase in IL-6 was observed in poled group (p>0.05). Fibroblasts secretion of IL-1β was similar to osteoblasts, with constant values over time in line with several studies with other cell lines in literature.⁷⁻⁸ However, IL-6 secretion decreased in all groups, with values significantly lower in non-poled group compared to YZS at 3 days (p>0.05), which suggests a possible anti-inflammatory role of non-poled BaTiO₃. The results of this study suggest that zirconia samples with 5% of BaTiO₃ are not cytotoxic to peri-implant tissue cells and, additionally, do not seem to affect their long-term inflammatory profile. In this sense, further studies should be carried out to assess the potential piezoelectric effect on bone tissue when load is applied and in a biological systems.

CONCLUSION

Zirconia composite surfaces with the addition of BaTiO₃ is not cytotoxic to peri-implant tissues cells. Additionally, samples with or without piezoelectric properties do not affect cellular differentiation and inflammatory profile. Nevertheless, the addition of non-poled BaTiO₃ to YSZ may have a potential reduction effect in IL-6 mediated —inflammatory activity in fibroblasts.

ACKNOWLEDGMENT

This work was supported by FCT (Foundation for Science and Technology - Portugal) under the FunImp project 01-0145-FEDER-030498. Co-financed by:



REFERENCES

- Cionca N, Hashim D, Mombelli A. Zirconia dental implants: where are we now, and where are we heading? *Periodontol* 2000. 2017;73(1):241-58.
- Hirano T, Sasaki H, Honma S, Furuya Y, Miura T, Yajima Y, et al. Proliferation and osteogenic differentiation of human mesenchymal stem cells on zirconia and titanium with different surface topography. *Dent Mater J*. 2015;34(6):872-80.
- Furuya K, Morita Y, Tanaka K, Katayama T, Nakamachi E. Acceleration of osteogenesis by using barium titanate piezoelectric ceramic as an implant material. *Proc. SPIE* 7975. *Bioinspiration, Biomimetics, and Bioreplication*. 2011; 79750U
- Rocca A, Marino A, Rocca V, Moscato S, de Vito G, Piazza V, Mazzolai B, Mattoli V, Ngo-Anh TJ, Ciofani G. Barium titanate nanoparticles and hypergravity stimulation improve differentiation of mesenchymal stem cells into Osteoblasts. *Int J Nanomedicine*. 2015;8(10):433-45.
- Fan B, Guo Z, Li X, Li S, Gao P, Xiao X, Wu J, Shen C, Jiao Y, Hou W. Electroactive barium titanate coated titanium scaffold improves osteogenesis and osseointegration with low-intensity pulsed ultrasound for large segmental bone defects. *Bioact Mater*. 2020 Jul 15;5(4):1087-1101.
- Tainen L, Gasik M, Pinto J, Carvalho O, Silva F. Thermal Stability of BaTiO₃ Functionalised Zirconia. *eCM Online Periodical*, 2019, Collection 2; ScSB Conference Abstracts (page 63)
- Majumdar S, Hira SK, Tripathi H, Kumar AS, Manna PP, Singh SP, Krishnamurthy S. Synthesis and characterization of barium-doped bioactive glass with potential anti-inflammatory activity. *Ceramics International*. 2021; 47(5):7143-7158.
- Ball JP, Mound BA, Nino JC, Allen JB. Biocompatible evaluation of barium titanate foamed ceramic structures for orthopedic applications. *J Biomed Mater Res A*. 2014 Jul;102(7):2089-95